

Prevalence of cutaneous myiasis along with secondary bacterial complications in ruminants

Supta Nusrat Jahan¹, Juli Mst Sogra Banu^{*1}, Hoque Md Fazlul¹, Kibria Md Golam², and Islam Md Nazrul³

ABSTRACT

Myiasis is a common and widespread clinical problem closely interlinked with serious economic losses of livestock in Bangladesh, though it is thought to be a neglected cutaneous infection. The study was carried out to determine the clinical prevalence of myiasis in cattle, sheep and goat based on different epidemiological parameters such as species, sex, age and season with special emphasis to identify the secondary bacterial complications in maggot wounded areas. The swabs were randomly collected from the myiatic wounds, brought to Microbiological laboratory with necessary cautions followed by bacteriological cultivation, isolation, and identification. The data were recorded and organized systematically and analyzed statistically. A total number of 4149 clinical cases were recorded, where 312 cases were registered as myiasis irrespective of species. The overall prevalence of myiasis was found at 7.52%. Higher prevalence of myiasis were noticed in cattle (11.67%) (n=187) followed by goat (5.01%) (n=122) and sheep (2.68%) (n=3) that differed significantly (P<0.05). Myiasis showed higher in female animals (22.39%) than in males (14.92%), and young animals (52.41%) were predominantly susceptible than adults (6.42%) in cattle and goats whereas the opposite scenario was seen in sheep. Seasonally the highest clinical cases were found in summer (42.62%) followed by rainy (15.00%) and winter season (2.92%) which demonstrated statistically significant (P<0.05) in cattle and goat but insignificant (P>0.05) in case of sheep. In myiatic wound, the bacterial complications were higher with *Staphylococcal spp* infection (72.22%) followed by *Escherichia coli* (11.11%). Combined infection of *E. coli* and *Klebsiella spp.* was 11.11%, whereas single *Klebsiella* infection was 5.56%.

Key Words: Bacteria, myiasis, prevalence, ruminants.

INTRODUCTION

Myiasis in domestic animals is a worldwide problem including Bangladesh causing serious economic losses in livestock through reduced productivity (milk, meat, wool) and tissue damage (poor quality hide). It impairs livestock production not only by inducing mechanical damage to internal organs and skin but also by downregulating the immune system of the host (Boulard, 2002).

Myiasis is the invasion of tissues or organs of live vertebrates (humans and/or animals) with dipterous larvae, which feed on host's dead or living tissues, liquid body substances or ingested foods (Zumpt, 1965). The most frequent host for myiasis is cattle (46.4%) followed by dogs (15.3%), humans (14.7%), pigs (6%), horses (4%) and sheep (1%) (Sergio et al., 2007). Wounds, ulcers, sores,

broken horns, injured eyes, and disrupted skins are the commonest predisposing factors of myiasis in domestic animals (Juyena et al., 2013). The female fly lays eggs on new wounds followed by the hatching of larvae that creep, feed, and live in the tissue (Hamid, 2016). Oviposition is encouraged by a foul-smelling and purulent discharge from diseased tissue. The exact chemosensory causing attraction for oviposition is unknown, but the presence of pus and bacterial metabolites in the wounds are known to increase the attractiveness of wounds as oviposition sites (Humphery et al., 1980).

The severity of the disease is often become fatal due to secondary bacterial complication followed by bacteremia, septicemia or toxemia (Guerrini et al., 1988; Schnur et al., 2009). Among gram-positive aerobes, *Staphylococcus aureus*, *Staphylococcus albus*, and *Streptococcus pyogenes* were more frequently isolated from myiatic wounds. Gram-negative aerobes included *Proteus spp.* and *Escherichia coli*. (Malaviya, 2005; Juyena et al., 2013). Therefore, the present research

work was undertaken to clinically observe the prevalence of myiasis in livestock species along with the isolation of invading secondary bacteria from the affected myiatic wounds.

MATERIALS AND METHODS

The study was conducted for a year ranging from May 2017 to April 2018. A total of

The visible clinical manifestations of the affected animals were noted systematically and clinical cases were recorded during the physical visit of the farms and the farmer's complaints were also emphasized. The epidemiological data were recorded based on species of animals affected, sex, age and season, and groupings were done based on the factors as stated above (Table 1).

Table 1: Overall prevalence of myiasis in ruminants

Total clinical cases	Myiasis affected cases	Prevalence (%)
4,149	312	7.52

Table 2: Species-wise prevalence of myiasis in ruminant animals

Species	Total No. of Clinical cases	Myiasis affected cases	Prevalence (%)	P-value
Cattle	1602	187	11.67	0.00 (**)
Goat	2435	122	5.01	
Sheep	112	3	2.68	

* means 5% level of significant ($p < 0.05$)

** means 1% level of significant ($p < 0.01$)

NS means non-significant ($p > 0.05$)

Table 3: Frequency distribution of myiasis according to species in ruminants

Species	Population affected with myiasis	No. of individuals affected with myiasis	Percentage (%)
Cattle	(n=312)	187	59.94
Goat		122	39.10
Sheep		03	0.96

Table 4: Sex-wise prevalence of myiasis in ruminants

Species	Male			Female			P- value
	Total No. of cases	Affected with myiasis	Prevalence %	Total No. of cases	Affected with myiasis	Prevalence %	
Cattle	572	53	9.27	1030	134	13.00	0.02(*)
Goat	1136	40	3.52	1299	82	6.31	0.00 (**)
Sheep	47	1	2.13	65	2	3.08	0.76(NS)

Table 5: Frequency distribution of myiasis according to Sex in ruminants

Sex of animal	Cattle		Goat		Sheep	
	Total affected	Percentage (%)	Total affected	Percentage (%)	Total affected	Percentage (%)
Male	53	28.34	40	32.79	1	33.33
Female	134	71.66	82	67.21	2	66.67

4149 clinical cases were recorded among which only 312 myiasis affected animals (cattle, goat, and sheep) were registered during the experimentation.

Microbiological study

The affected animals were randomly selected (at least 3 in each species) to collect swabs, swabs were brought to bacteriology laboratory (Department of Microbiology,

Hajee Mohammad Danesh Science and Technology University, Dinajpur) with necessary cautions, attempted to cultivate

RESULTS AND DISCUSSION

A total number of 312 myiasis affected cases

Table 6: Frequency distribution of myiasis according to age in cattle

Age of cattle	Affected with myiasis	Percentage (%)	P value
Group-1 (0-1 year)	98	52.41	0.00 (**)
Group-2 (>1to 2years)	36	19.25	
Group-3 (>2 to 4 years)	41	21.93	
Group-4 (>4 years)	12	6.42	

Table 7: Frequency distribution of myiasis according to age in sheep and goat

Age	Goat		Sheep	
	Affected with myiasis	Percentage (%)	Affected with myiasis	Percentage (%)
Group-1 (0-6 months)	22	18.03	1	33.33
Group-2 (>6 to 12 months)	25	20.49	-	-
Group-3 (>12 to 24 months)	33	27.05	2	66.67
Group-4 (>24 months)	42	34.43	-	-
Total	122	100	3	100
P-value	0.01 (*)		0.41 (NS)	

Table 8: Season-wise prevalence of myiasis in ruminants

Species	Summer Season n=1208			Rainy Season n=1444			Winter Season n=1497			P-value
	No. of cases	Affected with myiasis	Prevalence %	No. of cases	Affected with myiasis	Prevalence %	Total no. of cases	Affected with myiasis	Prevalence %	
Cattle	503	120	23.86	498	55	11.04	601	12	2.00	0.00 (**)
Goat	662	78	11.78	908	36	3.96	865	8	0.92	0.00 (**)
Sheep	43	3	6.98	38	-	-	31	-	-	0.08 (NS)

organisms in solid and liquid media, isolation and identification were done based on the classical growth, staining and morphological properties of the bacteria as per standard procedures (Merchant and Packer 1967; Cheesbrough, 1985)

Statistical analysis

The clinical data were noted and organized systematically and analyzed statistically by using software 'SPSS' (version 17) following chi-square (χ^2) test

were identified in cattle, goat, and sheep from 4,149 different clinical cases at Sadar Upazila of Dinajpur district for a period of one year. In this study, the overall prevalence was 7.52% (Table 1) which is comparable to Samaddar et al. (2016) and Imtiaz et al. (2014) who reported 10.97% out of 5987 in Coastal region of Bangladesh, 5.21% out of 4338 in Chittagong, Bangladesh respectively. Sarker et al. (2016), Juli et al. (2015), Giangaspero et al. (2011), Alahmed (2004) who reported the overall

prevalence of myiasis was 4.09% at Chittagong, 4.11% at Dinajpur of Bangladesh, 3% in Italy, 2% in Riyadh respectively. The higher prevalence rate than the present study was reported that are 13.4% in Bathans of Pabna and Sirajgonj

Among the three species, the occurrence of myiasis in cattle was 59.94%, goat 39.10% and sheep 0.96% which is shown in Table: 3. Juyena et al. (2013) and Sergio et al., (2007) stated that cattle as the most frequent hosts for myiasis and their results are

Table 9: Frequency distribution of myiasis according to season

Seasons	Cattle		Goat		Sheep	
	No. of affected animal	Percentage (%)	No. of affected animal	Percentage (%)	No. of affected animal	Percentage (%)
Summer season	120	64.17	78	63.93	3.0	100
Rainy Season	55	29.41	36	29.51	0.	0.0
Winter season	12	6.42	8	6.56	0.0	0.0
Total	187	100	122	100	3	100

Table 10: Percentage of isolated bacteria from myiasis affected wounds

List of organisms	Total No. of organisms	Percentage (%)
<i>Staphylococcus spp.</i>	13	72.22
<i>Escherichia coli</i>	2	11.11
<i>Klebsiella spp.</i>	1	5.56
<i>Escherichia coli</i> + <i>Klebsiella spp.</i>	2	11.11
Total	18	100

districts (Sarker et al., 2013), 14.71% in southeastern part of Iran (Radfar and Haji Mohammad, 2012), 13.1% in South Iran (Shoorijeh et al., 2011), 59.9% in Ethiopia (Gebremedhin, 2011), 40.3% in north-eastern part of Turkey (Arslan et al., 2008), 31.9% in Turkey (Kara et al., 2005), 24% in northern Jordan (Abo-Shehada et al., 2003) and 35.68% cases in northern Mediterranean region of France (Dorchies et al., 2000).

Species wise prevalence of myiasis

During the study, the prevalence of myiasis in cattle was 11.67%, goat 5.01% and sheep 2.68% which was highly significant (Table: 2). The findings are more or less similar to Samaddar et al., (2016) who stated cattle 6.16%, goat 5.42%, and sheep 0.016%. Yadav et al., (2013) found the prevalence rate of myiasis 9.73% out of 750 cattle in the Jammu region, India.

correlated with the result of the present study. The prevalence of myiasis in cattle was 3.56–10.23% in Turkey (Gulanber et al., 2000), 8% in Canada (Colwell, 2000) which is similar to our study. This result is in contrast with the reports of Imtiaz et al., (2014) who investigated that the highest occurrence was found in goat than other animals. The prevalence of myiasis in cattle was 1.67% and goat 2.42% in Chittagong and Cox’S Bazaar district of Bangladesh (Sarker et al., 2016) which is also dissimilar to our study. The higher prevalence rate than the present study was reported in cattle 23.80% in Manikgonj (Rahman et al., 2013), 24.7% in Babugonj, Barisal (Rahman et al., 2012), 35.50% in Chakwal, Punjab, Pakistan (Hasan et al., 2008), 85% in Italy (Frangipane et al. 2003), and 80% in China (Yin et al. 2003) those are comparable to our result.

Sex- wise prevalence of myiasis

The study showed that female animals were significantly ($p \leq 0.05$) higher infested with myiasis than males. Among three species (cattle, goat, sheep) prevalence of myiasis in female animals (Overall= 22.39%) were

Juyena et al., (2013) who reported that the infestation of myiasis was higher in females (64.5% and 66%) than male (35.5% and 34%). Samaddar et al., (2016), Alejandro et al., (2015) and (Radfar and Haji Mohammadi, 2012) also reported that the

Table 11: Cultural and Morphological Characteristics of Isolated Bacteria

Name of bacteria	Culture in media	Cultural Characteristics	Staining and Morphological Characteristics
<i>Staphylococcus</i> spp.	Nutrient agar	Grayish white to yellowish colonies	Gram positive cocci, violet color arranged in grape like cluster
	MacConkey Agar	Pale color / colorless colonies	
	Mannitol salt agar	Golden yellow color colonies	
<i>Escherichia coli</i>	Nutrient agar	Gray white colonies	Gram negative, pink color large rod shape appearance, arranged in single or paired
	MacConkey Agar	Lactose positive, Rose pink colour colonies	
	Eosin Methylene Blue (EMB) agar	Smooth, circular, small green colonies with dark center and metallic sheen	
<i>Klebsiella</i> spp.	Nutrient agar	Grayish colonies	Gram negative, pink color, small rod shaped organism
	MacConkey Agar	Mucoid, convex, lactose positive, pink color colonies	
	Eosin Methylene Blue (EMB) agar	Large, mucoid, pink to purple colour colonies without metallic	

Table 12: Identification of Isolated Bacteria by Biochemical Tests

Name of bacteria	Indole test		MR test		VP test		Catalase test	
	Change	Result	Change	Result	Change	Result	Change	Result
<i>Staphylococcus</i> spp.	No color change	-(ve)	Red color	+(ve)	Red color	+(ve)	Bubble formation	+(ve)
<i>Escherichia coli</i>	Pink color ring at the top of the media	+(ve)	Red color	+(ve)	No color change	-(ve)	Bubble formation	+(ve)
<i>Klebsiella</i> spp.	No color change	-(ve)	No color change	-(ve)	Red color	+(ve)	Bubble formation	+(ve)

13.00%, 6.31% and 3.08% where male animal (Overall 14.92%) were 9.27%, 3.52% and 2.13%, respectively (Table: 4). There was an insignificant sex-wise variation found in sheep.

The frequency of myiasis in female animals in cattle (71.66%) and in goat (67.21%) were higher than males (28.34%; 32.79%), respectively (Table: 5). These findings have a similarity with Imtiaz et al., (2014) and

affection rate of myiasis is higher in female than male. These results support the earlier investigations of Sarker et al., 2014 (11.6% in male and 14.0% in female) in the case of cattle. Ozdal et al., (2016) and Osman et al., (2017) said that the infestation rates were 41.26 % and 13.9% in female and 34.42 % and 2.2% in male respectively. The findings of Sarker et al., (2016), Islam et al., (2015) and Rahman et al., (2009) were similar to

these findings. Saleem et al., (2017) and Hasan et al., (2008) stated that the prevalence was higher in males (100 %; 39.50%) compared to females (98.2%; 31.50%) animals respectively which is a contrast to this findings. *Negm-Eldin* et al.,

(Table: 6) which is close to *Juyena* et al., (2013) stating the occurrence of myiasis was higher (41%) in animals of less than 6 months of age. *Juli* et al., (2015) and *Hasan* et al., (2008) stated that the occurrence of myiasis was higher in young animals than

Clinical Findings

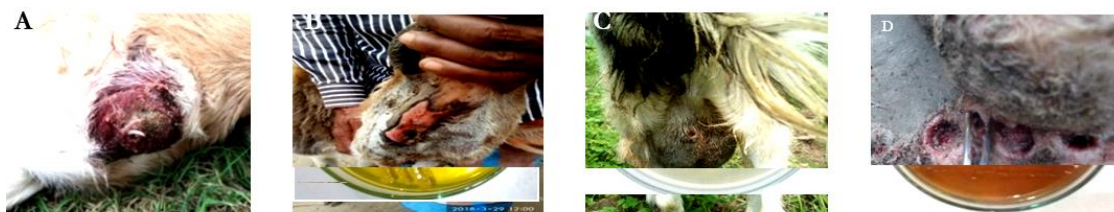


Fig. 1: A. Myiasis in naval region, B. head region of cattle, C. scrotal region of buck D. Inguinal region of calf

Cultural and morphological characteristics of *Staphylococcus Spp.*

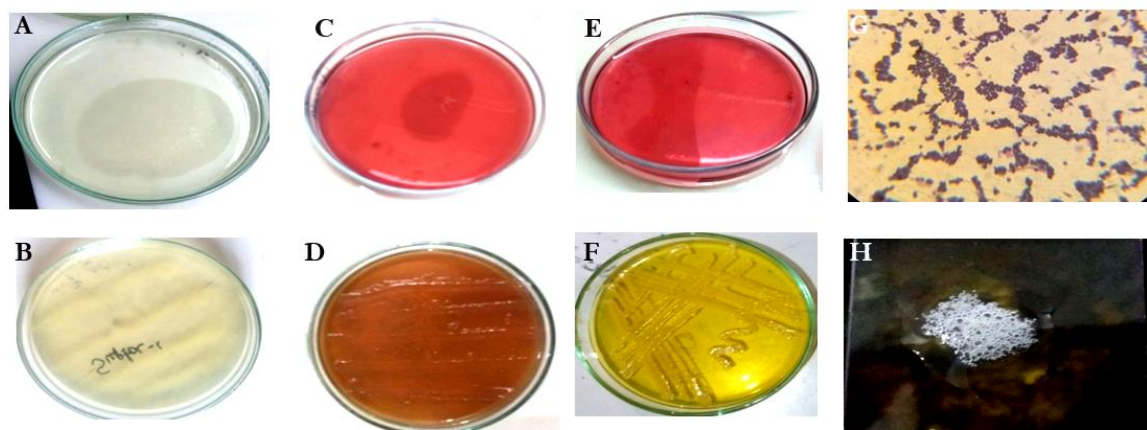


Fig 2:A. Nutrient Agar (Control media), B. Produce gray white to yellowish color colonies in Nutrient Agar media, C. MacConkey agar (control media), D. Pale color colonies of *Staphylococcus spp* in MacConkey agar, E. Mannitol Salt Agar (control media) F. Golden yellowish color colonies in MSA, G. Morphological characteristics of *Staphylococcus Spp.* showing violet color cocci, arranged in grape like cluster in gram staining method. H. Catalase positive.

(2015); *Mot* (2013); *Orfanou* et al. (2011), *Abd El- Rahman* (2010); *Kara* et al. (2005) reported more cases in male than the female which is also dissimilar to this study. *Oryan* et al. (2012), *Shoorijeh* et al. (2011), *Tavassoli* et al. (2010) and *Abo- Shehada* et al. (2003) found the same type of infection rate in both sexes which is a contrast to our findings.

Age-wise prevalence of myiasis

The infection rate was significantly higher in cattle less than one year of age (0-1year) 52.41% than the older age group (>4 years)

the adult animals which is similar to our study. *Kara* et al. (2005) said the intensity of the infestation decreased with the age of cattle which is parallel to the present study. This result is in contrast with the reports of *Islam* et al. (2015) and *Rahman* et al. (2009) who investigated that the highest occurrence 44% and 40% was found in the adult animals of over 2 years and the lowest 25% and 27% in the calves of below 6 months respectively. *Yadav* et al., (2013) found that the highest (13.57%) prevalence occurred in 1-4 years of age and lowest (0.68%) is below

1 year of age groups of the animal which is dissimilar to our study. Dehghani et al. (2012) reported that the prevalence of myiasis was higher in the old age group than young which not supported by this study. An infestation of myiasis in sheep due to

(34.43%) and the lowest infection in young animals below 6 months (18.03%) in case goat (Table: 7) which is comparable to Arslan et al. (2008) stating that the infestation rate up to 1-years-old was 30.0%, 1 to 3 years-old 40.0% and older than 3

Cultural and morphological characteristics of *E. coli*

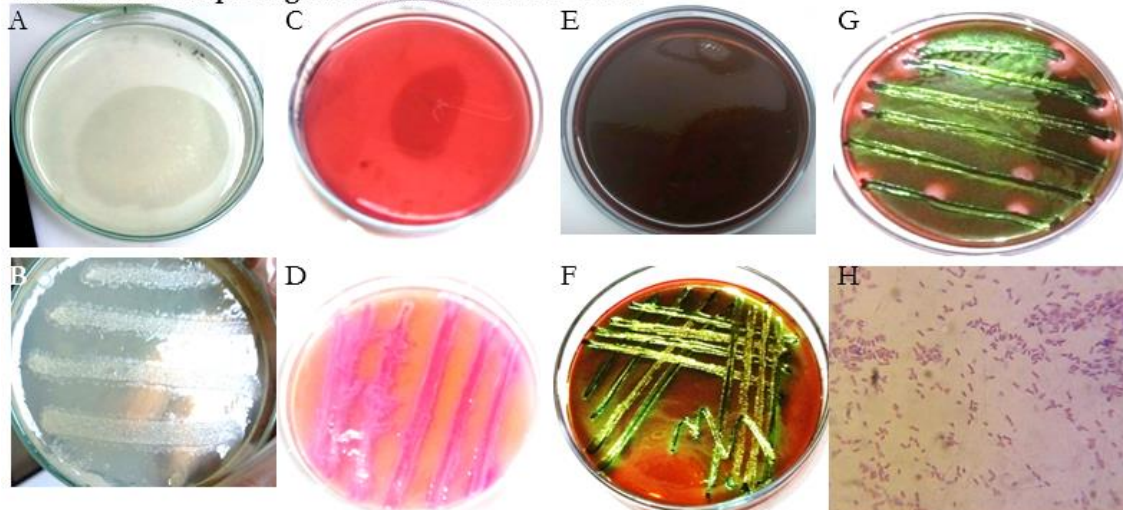


Fig. 3: A. Nutrient Agar (Control media), B. Pale color colonies in Nutrient agar media, C. MacConkey agar (control media), D. Rose pink colour colonies in MacConkey agar, E. EMB agar (control media), F. Green metallic sheen in EMB agar, G. Green metallic sheen with pink color colonies in *E. coli*+ *Klebsiella* spp. in EMB agar H. Microscopic morphology of *Escherichia coli* showing pink color large rod shaped bacteria

Cultural characteristics of *Klebsiella* spp.

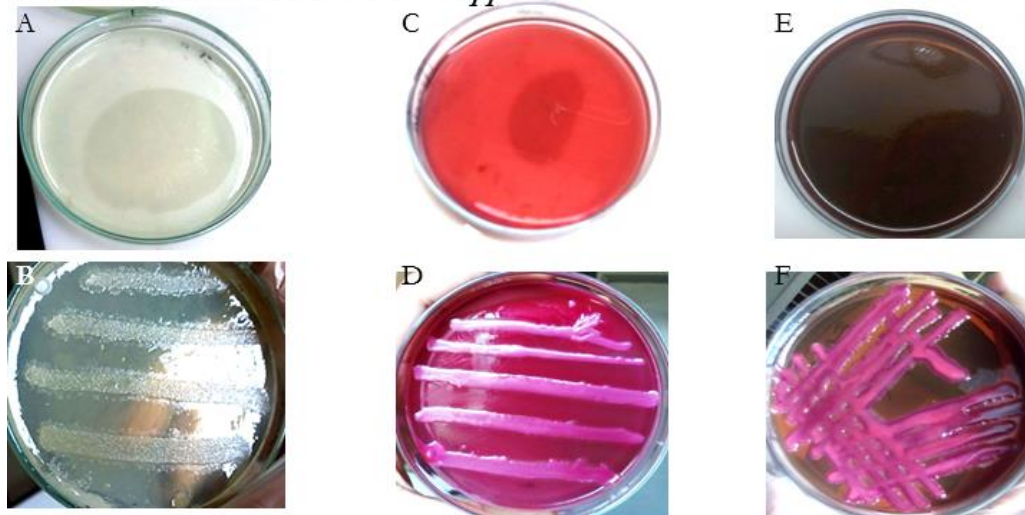


Fig.4: A. Nutrient Agar (Control media), B. Pale color colonies in Nutrient agar, C. MacConkey agar (control media), D. Mucoid, convex, pink color colonies in MacConkey agar, E. EMB agar (control media), F. Large, mucoid, pink to purple colour colonies in EMB agar

age appeared insignificant ($P>0.05$). The study indicates that the highest infection was found in over 24 months

years old was 52.4% in small ruminants. Shoorijeh et al. (2011) stated that infestation rates were significantly higher in 4-5-year-

old goats (16.2%) than 6 months to 2-year-old (7.4%) groups. *Negm-Eldin et al. (2015)* and *Tavassoli et al. (2010)* said that the prevalence was significantly higher in sheep and goat of 25-36 months old than bellow 6 months of age which is close to our present

and *Genet, 2000; Scala et al., 2001; Alem et al., 2010; Karatepe et al., 2014*) which is a contrast to this result. *Saleem et al. (2017)* reported that all age groups of animals were equally susceptible to infestations.

Season- wise prevalence of myiasis

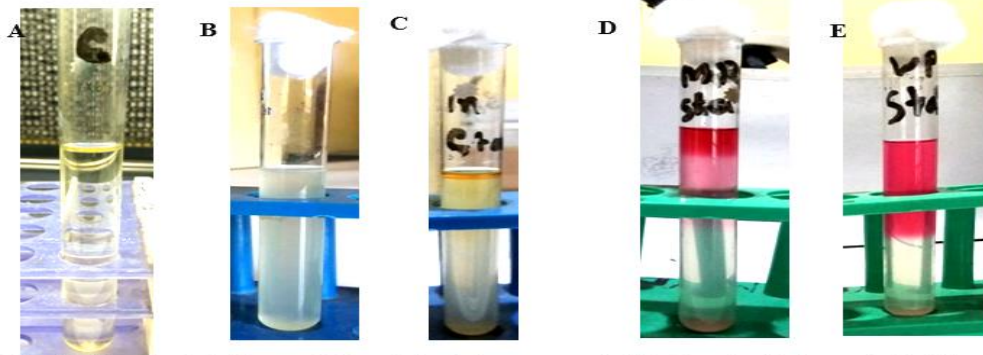


Fig. 5: Biochemical characteristics of *Staphylococcus* spp. A. Nutrient broth (control) B. Pale color colonies in Nutrient broth C. Indole (-ve), D. MR (+ve), E. VP (+)

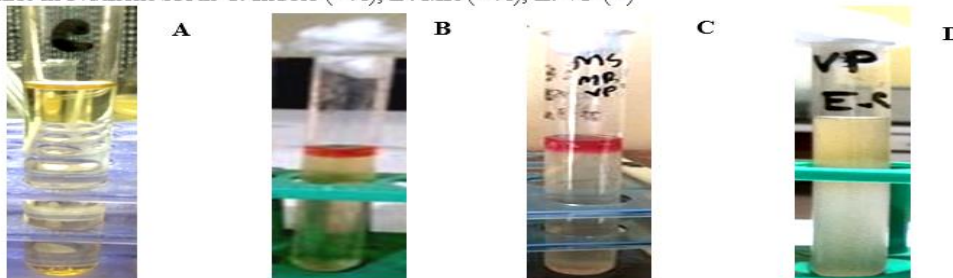


Fig. 6: Biochemical characteristics of *E. Coli*. A. Control B. Indole (+ve), C. MR (+ve), D. VP (-ve)

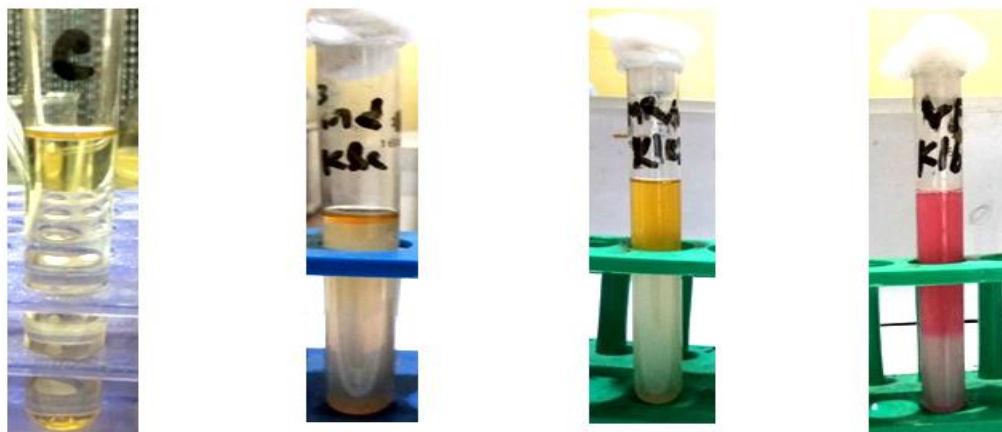


Fig. 7: Biochemical characteristics of *Klebsiella* spp. A. Control Media B. Indole (-ve), C. Control Media, D. MR (-ve), E. VP (+)

study. This finding was consistent with that was recorded by *Abo-Shehada et al. (2000)*, *Uslu and Dik (2008)* who reported higher infestation rates above 3 years. In goat, Many authors reported a significantly higher infestation rate in young than adult (*Yilma*

The study showed that the highest prevalence of myiasis in cattle, goat and sheep were found in summer season (overall, 42.62%); (23.86%, 11.78% and 6.98%) which was followed by rainy season (overall, 15%); (11.04%, 3.96%, 0%) and

winter season (overall, 2.92%); (2.00% & 0.92%, 0%) respectively (Table: 8). There was insignificant variation found in sheep in occurrences of myiasis according to season. Occurrence of myiasis in cattle and goat were 64.17% & 63.93% in summer season, 29.41% & 29.51% in rainy season, 6.42% & 6.56% in winter season, respectively were recorded (Table: 9). Rahman et al., (2012) and Silva et al., (2012) recorded the highest myiasis cases in cattle and goats during the summer season in comparison to the winter season which closes to this result. Juli et al., (2015) and Sarker et al. (2014) investigated the higher rate of myiasis in summer than winter and moderate in the rainy season. These findings are inconsistent with that of Juyena et al. (2013) and they stated higher incidences of myiasis during March-June (summer season) 80% followed by July-October (rainy season) 13% and November-February (winter season) 7%. Alahmed, (2004) found a higher incidence of myiasis during March-May (60%) due to optimum temperature and relative humidity. He also reported lower infestation during the dry hot season (June-August, 5%) and cold season (December-February, 1.5%) and his results more or less similarly correlated with the present study. Ipek, (2017) observed seasonal prevalence in goats varied between 44.18% and 100%, with the highest prevalence being in the winter and the lowest prevalence in the autumn which is not parallel to this result. Osman et al. (2017), Bosly (2013) and Abd El-Rahman (2010) stated the rate of infestation was significantly greater in the colder months compared to those of warmer months. Hasan et al. (2008) said, bovine myiasis was found only from September to December. In contrast, winter peak larval infestations were observed by Alcaide et al. (2003) and Shoorijeh et al. (2009).

Isolation and identification of bacteria

During this study, the highest prevalence of *Staphylococcus* spp. 72.22% was seen followed by *E. coli* 11.11%, combined infection by *E. coli* + *Klebsiella* spp. 11.11%

and single *Klebsiella* spp. infection 5.56% (Table: 10). The cultural, morphological and biochemical characteristics of *Staphylococcus* spp (Fig.: 2 & Fig: 5), *E.coli* (Fig.: 3 & Fig.: 6) and *Klebsiella* spp (Fig.: 4 & Fig.: 7) are listed in Table: 11 & Table: 12. Islam et al. (2015) and Gunalan et al. (2011) reported that myiasis wounds are associated with *Staphylococcus* spp. Mot, (2013) detected bacteria from maggot wound samples where: *Staphylococcus aureus*, *Bacillus subtilis*, *Proteus vulgaris*, *Micrococcus luteus* and *Escherichia coli* were isolated which close to the present findings. El-Nouby et al. (2009) identified the following pathogenic bacteria species from myiatic wounds: coagulase-negative *Staphylococci*, including methicillin-resistant *Staphylococcus aureus* (MRSA), *Bacteroides*, *Proteus*, *Pseudomonas* and *Klebsiella* spp. Malaviya, (2005) reported that the bacterial flora seen in maggot-infested wounds of the extremities. Among gram-positive aerobes, *Staphylococcus aureus*, *Staphylococcus albus*, and *Streptococcus pyogenes* were more frequently isolated and gram-negative aerobes included *Proteus* spp. and *Escherichia coli*. Anaerobes which were isolated include *Micrococcus* and *Bacteroids* whereas Clostridia were seen infrequently (2% cases).

CONCLUSION

Myiasis is the most familiar problem in veterinary practice. A cross-sectional study was carried out to investigate the prevalence of myiasis in domestic animals according to species, age, sex and season. Samples were collected for isolation and identification of major pathogenic bacteria from maggot affected wounds. The overall prevalence of myiasis was 7.52% in livestock animals (cattle, goat, and sheep). Cattle are the most susceptible species for myiasis (11.67%) than the goat (5.10%) and sheep (2.68%) in this region. Sex, age and season influenced the prevalence of myiasis in domestic animals. The major pathogens isolated from maggot wounds were *Staphylococcus* spp.

, *Escherichia coli*, and *Klebsiella* spp. which increases the attractiveness of Diptera flies. These findings could create awareness in animal rearers by generating the basic data which may help to prevent maggot infestation and control the spread of myiasis

ACKNOWLEDGEMENT

The authors are highly grateful to the Veterinary Surgeon, Upazila Livestock Office, Sadar, Dinajpur, Bangladesh for assisting physical visit, data collection, necessary suggestions; Department of Microbiology, Hajee Mohammad Danesh Science and Technology University, Dinajpur, Bangladesh for cordial cooperation during bacteriological analysis; and Ministry of Science and Technology, Bangladesh for funding in this research work.

REFERENCES

1. Abd El-Rahman SS (2010). Prevalence and Pathology of Nasal Myiasis in Camels Slaughtered in El-Zawia Province-Western Libya: with a Reference to Thyroid Alteration and Renal Lipidosis. *Global Veterinaria*; 4(2): 190-197.
2. Abo-Shehada MN, Arab B, Mekbel R, Williams D, Torgeson PR (2000). Age and seasonal variations in the prevalence of *O. ovis* larvae among sheep in the northern Jordan. *Preventive Veterinary Medicine*; 47: 205-212.
3. Abo-Shehada MN, Batainah T, Abuharfeil N, Torgeson PR (2003). *Oestrus ovis* larval myiasis among goats in northern Jordan. *Preventive Veterinary Medicine*; 59: 13-19.
4. Acha PN, Szyfres B (2003). Zoonosis by Enfermedades Transmisibles Comunesal Hombrey los Animales, 3rd ed., OPS Pub. Cient. Tec. No. 80. pp. 412.
5. Alahmed AM (2004). Myiasis in sheep farms in Riyadh region, Saudi Arabia. *Journal of the Egyptian Society of Parasitology*; 34(1): 153-160.
6. Alcaide M, Reina D, Sanchez J, Frontera E, Navarrete I (2003). Seasonal variations in the larval burden distribution of *O. ovis* in sheep in the south west of Spain. *Veterinary Parasitology*; 118: 235-241.
7. Alejandro H, Héctor P, Carlos O, Flery FS (2015). *Oestrus ovis* infection of grazing sheep during summer in southern Chile. *Pesquisa Veterinária Brasileira*; 35(6): 497-500.
8. Alem F, Kumsa B, Degefu H (2010). *Oestrus ovis* larval myiasis among sheep and goats in Central Oromia, Ethiopia. *Tropical Animal Health and Production*; 42: 697-703.
9. Arslan MO, Kara MM, Gicik YY (2008). Epidemiology of *Oestrus ovis* infestations in sheep in Kars province of north-eastern Turkey. *Tropical Animal Health Production*; 41: 299-305.
10. Bosly HA (2013). Seasonal prevalence of *Oestrus ovis* (Diptera: Oestridae) larvae in infested sheep in Jazan Region, Saudi Arabia. *Journal of Parasitology and Vector Biology*; 5(5): 66-71.
11. Chhabra MB, Pathak KML (2009). Myiasis of domestic animals and man in India. *Journal of Veterinary Parasitology*; 23(1): 1-7.
12. Cheesbrough M. (1985). *Culturing of Anaerobs. International Medical Laboratory Manual for Topical Countries*. Butler Worth Co. Vent. 2: 50-56.
13. Colwell DD (2000). Persistence of cattle grub (Diptera: Oestridae) on a Canadian ranch with long term, continuous therapeutic control. *Veterinary Parasitology*; 94:127-132.
14. Dehghani R, Sedaghat MM, Esmaeli N, Ghasemi A (2012). Myiasis among slaughtered animals in Kashan, Iran: descriptive a veterinary entomological problem in the tropics. *Iranian Journal of Veterinary Science and Technology*; 4(1): 19-28.

15. Dorchies PH, Bergeaud JP, Tabouret G, Duranton C, Prevot F, Jacquet PH (2000). Prevalence and larval burden of *Oestrus ovis* (Linné 1761) in sheep and goats in the northern Mediterranean region of France. *Veterinary Parasitology*; 88: 269-273.
16. El-Nouby KA, El-Heniedy MA, Hassan AM (2009). Parasitological, Bacteriological and Clinical Study of Wound Myiasis with a Trial of Maggot Therapy in Intractable Wounds. *Egyptian Journal of Medical Microbiology*; 18(4): 77-88.
17. Frangipane di Regalbano A, Capelli G, Otranto D, Pitrobelli M (2003). Assessment of cattle grub (*Hypoderma* spp.) prevalence in northeastern Italy: an immune-epidemiological survey on bulk milk sample using ELISA. *Veterinary Parasitology*; 111: 343-350.
18. Gebremedhin EZ (2011). Prevalence of ovine and caprine oestrosis in Ambo, Ethiopia. *Tropical Animal Health Production*; 43(1): 265-270.
19. Gingaspero A, Traversa D, Trentini R, Scala A, Otranto D (2011). Traumatic myiasis by *Wohlfahr tiamagnifica* in Italy. *Veterinary Parasitology*; 175(1-2): 109-112.
20. Guerrini VH (1988). Ammonia toxicity and alkalosis in sheep infested by *Lucilia cuprina* larvae. *International Journal of Parasitology*; 18: 79-81.
21. Gulanber, A, Tuzer E, Gargili A, Toparlak M, Efill I, Keles V, Ulutas M (2000). A survey of hypo-dermatosis cattle slaughtered in Thrace (Trakya) Turkey. *Turkish Journal of Veterinary and Animal Science*; 24: 429-430.
22. Gunalan S, Kamaliah G, Wan S, Rozita AR, Rugayah M, Osman MA, Nabijah D, Shah A (2011). Sheep Oestrosis (*Oestrus Ovis*, Diptera: Oestridae) in Damara Crossbred Sheep. *Malaysian Journal of Veterinary Research*; 2(2): 41-49.
23. Hasan MU, Khan MN, Iqbal Z, Khan IA, Chaudhry SA, Sajid MS (2008). Surveillance of Cattle Hypo-dermatosis in District Chakwal, Pakistan. *International Journal of Agriculture & Biology*. 10(3): 337-339.
24. Hope FW (1840). On insects and their larvae occasionally found in the human body. *Transactions of the Royal Entomological Society of London*; 256-271.
25. Humphery JD, Spradbery JP, Tozer RS (1980). *Chrysomya bezziana*; Pathology of Old World Screw-worm Fly Infestation in cattle. *Experimental Parasitology*; 49: 381-397.
26. Imtiaz MA, Rahman MA, Islam K, Barua M, Alim MA, Chowdhury S, Sikder S (2014). Prevalence and Associated Risk Factors of Myiasis in Different Areas of Chittagong, Bangladesh. *Research Journal for Veterinary Practitioners*; 2(2): 22-27.
27. Ipek DNS (2017). Prevalence and intensity of *Oestrus ovis* in sheep and goats in south-eastern part of Turkey. *Indian Journal of Animal Research, Agricultural Research Communication Centre*; doi: 10.18805/ijar.B-717: 1-6
28. Islam MT, Maruf AA, Mannan MA, Rahman HMR, Tarafder MM, Samad MA, Noman AA, Hossain MB, Rahman MM (2015). Isolation and identification of associated bacteria and maggots from myiasis affected wounds of cattle and goats in Bangladesh. *Journal of Advanced Veterinary and Animal Research*; 2(2): 95-100.
29. Juli MSB, Hoque MF, Badruzzaman ATM, Hossain MK (2015). Bovine diseases at Dinajpur district of Bangladesh: Epidemiological status with relation to age and season. *Annals of Veterinary and Animal Science*; 2(3): 55-63.
30. Juyena NS, Tapon MAH, Ferdousy RN, Paul S, Alam MM (2013). Retrospective study on occurrence of Myiasis in ruminants. *Progressive Agriculture*; 24 (1 & 2): 101 - 106.

31. Kara M, Arslan MO, Giclk Y (2005). The prevalence of bovine hypodermosis in Kars Province, Turkey. *Tropical Animal Health and Production*; 37(8): 617-622.
32. Karatepe B, Karatepe M, Güler S (2014). Epidemiology of *O. ovis* Larvae infestation in sheep in Nigde province, Turkey. *Revue De Médecine Vétérinaire*; 165 (7-8): 225-230.
33. Merchant IA, Packer RA. (1967). *Veterinary Bacteriology and Virology*. Seventh Edition. The Iowa University Press, Ames, Iowa, USA. pp. 286-306.
34. Mot D (2013). The Prevalence of Sheep Traumatic Myiasis in Western Romania and Bacteria Isolated from the Insects Maggots. *Scientific Papers: Animal Science and Biotechnologies*; 46(2): 437-440.
35. Negm-Eldin MM, Elmadawy RS, Hanan GM (2015). *Oestrus ovis* larval infestation among sheep and goats of Green Mountain areas in Libya. **Journal of Advanced Veterinary and Animal Research**. 2(4): 382-387.
36. Orfanou DC, Papadopoulos E, Cripps PJ, Athanasiou LV, Fthenakis GC (2011). Myiasis in a dog shelter in Greece: epidemiological and clinical features and therapeutic considerations. *Veterinary Parasitology*; 181(2-4): 374-378.
37. Oryan A, Bahrami S (2012). Pathology of natural *Przhevalskiana Silenus* infestation in goats. *Tropical Biomedicine*; 29(4): 524-531.
38. Osman SA, Omar HM (2017). Clinical and epidemiological studies on screwworm infestation in Qassim region, Saudi Arabia. *Tropical Biomedicine*; 34(4): 936-943.
39. Ozdal N, Tanritanir P, Ilhan F, Deger S (2016). The prevalence of ovine oestrosis (*Oestrus ovis* Linnaeus, 1761, Diptera: Oestridae) and risk factors in Eastern Turkey. *Veterinary Archive*; 86(3): 323-333.
40. Radfar MH, Hajmohammadi V (2012). Prevalence of goat warble fly, *Przhevalskiana silenus* in South-eastern Iran. **Scientia Parasitologica**; 13(2): 73-76.
41. Rahman MA, Hossain MA, Alam MR (2009). Clinical Evaluation of Different Treatment Regimes for Management of Myiasis in Cattle. *Bangladesh Journal of Veterinary Medicine*; 7(2): 348 -352.
42. Rahman MA, Islam MA, Rahman MA, Talukder AK, Parvin MS, Islam MT (2012). Clinical Diseases of Ruminants Recorded at the Patuakhali Science and Technology University Veterinary Clinic. *Bangladesh Journal of veterinary medicine*; 10(1&2): 63-73.
43. Rahman ML, Chowdhury MK, Hossain MSA, Shamsuddin M, Bhuiyan MMU (2013). Occurrence of Reproductive Diseases of Cattle at Saturia, Manikgonj. **Bangladesh Journal of Veterinary Medicine**; 11(2): 121-125.
44. Saleem T, Katoch R, Yadav A, Mir IA, Godara R, Ahamed I (2017). Prevalence of *Ovine oestrosis* in Plain and Kandi Areas of Jammu, North India. *International Journal of Science, Environment and Technology*; 6 (2): 1117 -1122.
45. Samaddar K, Rahman MM, Haque Z, Mia MH, Kundu P, Rahman MA (2016). An overview of surgical diseases of food animals in the coastal areas of Bangladesh. *International Journal of Natural and Social Sciences*. 3(2): 46-59.
46. Sarker MS, Sen A, Ahaduzzaman M, Sayeed MA, Bupasha ZB, Ferdous J, Manzoor S, Uddin N (2016). Occurrence, Risk Assessment and Wound Healing Approach of Myiasis in Cattle and Goat in Chittagong, Bangladesh. *Research Journal for Veterinary Practitioners*; 4(4): 60-65.
47. Sarker NU, Rahman MM, Rana MS, Islam MT, Rima UK (2013). Prevalence of Surgical Diseases of Cattle in Stall-

- Fed and Free range Cattle in Bangladesh. The Bangladesh Veterinarian; 30 (2): 62-69.
48. Sarker NU, Samaddar K, Haq MM, Rahman MM (2014). Surgical affections of cattle in the milk-shed areas of Bangladesh. The Bangladesh Veterinarian; 31(1): 38-45.
 49. Scala A, Solinas G, Citterio CV, Kramer LH, Genchi C (2001). Sheep oesterosis (*O. ovis* Linne 1761, Diptera: Oesteridae) in Sardinia. Veterinary Parasitology; 102: 133-141.
 50. Schnur HJ, Zivotofsky D, Wilamowski A (2009). Myiasis in domestic animals in Israel. Veterinary Parasitology; 161: 352-355.
 51. Sergio EB, José DE, Angel BC, Franklin C, Janina S, Sabina B, Enrique M (2007). Incidence of myiasis in Panama during the eradication of *Cochliomyia hominivorax* (Coquerel 1858, Diptera: Calliphoridae) (2002-2005). The Memórias do Instituto Oswaldo Cruz; 102(6): 675-679.
 52. Shoorijeh SJ, Negahban S, Tamadon A, Behzadi MA (2009). Prevalence and intensity of *Oestrus ovis* in sheep of Shiraz, southern Iran. Tropical Animal Health and Production; 41: 1259–1262.
 53. Shoorijeh JS, Tamadon A, Negahban S, Behzadi MA (2011). Prevalence of *Oestrus ovis* in goats of Shiraz, southern Iran. Veterinarski Arhiv; 81(1): 43-49.
 54. Silva BFD, Bassetto CC, Amarante AFTD (2012). Epidemiology of *Oestrus ovis* (Diptera: Oestridae) in sheep in Botucatu, State of São Paulo. Revista Brasileira de Parasitologia Veterinaria; 21(4): 386-390.
 55. Tavassoli M, Tajik H, Yaghobzadeh-Khangahi R, Javadi S (2010). Prevalence of Goat Warble Fly, *Præhensalskiana* spp. (Diptera: Oestridae), in West Azarbaijan, Iran. Iranian Journal of Veterinary Science and Technology; 2(1): 33-38.
 56. Uslu U, Dik B (2008). Prevalence and intensity of *O. ovis* in Akkaraman sheep in the Konya region of Turkey. Medical Veterinary Entomology; 20: 347-349.
 57. Yadav A, Katoch R, Khajuria JK, Godara R, Agrawal R (2013). Prevalence of *Hypoderma lineatum* in cattle of Jammu region. Journal of Parasitic Diseases; 37(2): 196-198.